

## Using Progesterone and Prostaglandin F<sub>2α</sub> for Ewes Estrus Synchronization during Summer Season in Egypt

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**Abstract:** This study was carried out to investigate the effect of administration of progesterone (P4) and prostaglandin F<sub>2α</sub> (PGF<sub>2α</sub>) hormones on ewe estrus synchronization during summer season in Egypt. The study was performed on thirty four non-pregnant or lactating local ewes, 2 - 4 years old and 42.1 ± 1.8 kg average live body weight. Ewes were randomly divided into three groups as follows: natural estrus (control n = 10), intervaginal progesterone sponges (P4 = 12 ewes) for 12 days and two injections of PGF<sub>2α</sub> (12 ewes) 12 days apart. Parameters of estrus, conception and body weight of lambs were measured. Results showed that the percentage of estrus signs exhibition in PGF<sub>2α</sub> group reached 83.3%, while the lowest percentage (60%) was observed in control group (P<0.05). The time of starting estrus was earlier (P<0.05) in PGF<sub>2α</sub> and P4 groups (44.8 and 46h, respectively) than that in control group (58.4 h). The estrus duration was significantly longer (P<0.05) in PGF<sub>2α</sub> and P4 groups (44 and 41.6 h, respectively) than control group (31.2 h). The percentages of pregnant and lambing rates were 100% in all animals came to heat in all groups. While pregnancy rates and lambing rates according to initial number used were 83.3, 75% and 60% for PGF<sub>2α</sub>, P4 and control groups (P<0.05), respectively. The litter size, birth weight and weaning weight of lambs born did not differ significantly among groups. Weaning rate was the highest (P<0.05) in PGF<sub>2α</sub> group (92.9%) and the lowest in P4 group (85.7%). The percentage of ewes lambing twins was the highest (P<0.05) in P4 group (56%) and the lowest percentage in control group (33%). The percentage of male lambs was the highest (P<0.05) for ewes in PGF<sub>2α</sub> group (57.1%) and the lowest for ewes in control group (37.5%). In conclusion, PGF<sub>2α</sub> and P4 protocols could adequately induce estrus synchronization and improve some reproductive and productive traits in local ewes during summer season in Egypt. Further studies are recommended to proof results using more number of ewes.

**Keywords:** Estrus Synchronization, P4, PGF<sub>2α</sub>, ewes

### INTRODUCTION

Reproductive activity in sheep varies according to the seasonal changes (Gibson and Robinson, 1971). Seasonal changes in reproduction can be observed by changes in the proportion of females showing sexual receptivity (Knights *et al.*, 2002). Also, Huston (1983) reported that the twinning and fertility rate were increased in winter than summer. This necessitates applying a synchronization procedure to overcome this phenomenon. Estrus synchronization is a technique used to bring large number of animals in a flock into overt heat at the predetermined time (Panhwar, 2007). Hormonal treatment to control ovulation and reproduction is a prerequisite for successful breeding and increasing the number of pregnant females (Motlomelo *et al.*, 2002; Husein *et al.*, 2005), conception rate and poor fertility especially under semiarid conditions (Husein and Kridli, 2003). Progestagen-based protocols are commonly used worldwide (Abecia *et al.*, 2012). In numerous studies on small ruminants a high degree of estrus synchronization with the use of progestagen sponges (khalifa, 1993; Simonetti *et al.*, 2002; Dogan and Nur, 2006). ProstaglandinF<sub>2α</sub> is one of its analogues causes luteolysis in sheep having a functional corpus luteum at the time of treatment (khalifa, 1993; Turk *et al.*, 2008). Double PGF<sub>2α</sub> injections are common for estrus synchronization in ewes (khalifa, 1993; Ataman *et al.*, 2005; Zeleke *et al.*, 2005). Progestagen-based protocols have the potential for environmental contamination because of the residual P4 in devices and the addition of antibiotics to avoid vaginitis (Ataman and Akoz, 2006; Vinales *et al.*, 2011; Oliveira *et al.*,

2015). Moreover, the progestagen sponges are expensive. However, using PGF<sub>2α</sub> and/or its analogues are a good alternative, because they are rapidly metabolized in the lung and therefore, not accumulated in tissues (Davis *et al.*, 1980). Therefore, the present study aimed to investigate the effect of administration of P4 and PGF<sub>2α</sub> hormones on estrus synchronization in ewes during summer season in Egypt, and determining some reproductive and productive traits in estrus synchronized ewes.

### MATERIALS AND METHODS

This study was carried out at the Animal Production Farm in Ismailia Research Station, Animal Production Research Institute during summer season (July, 2019) (latitude 31°E and longitude 32°N). Thirty-four non-pregnant and non-lactating local ewes with 2-4 years old and 42.1±1.8 kg average live body weight were used. Ewes were housed in semi-open pens. The same feeding conditions (NRC, 1985) were applied to all animals. Animals were raised at the same environmental condition in pens. Ewes were randomly divided into three groups as follows: Control group (n=10): natural estrus occurring, P4 group (P4 = 12 ewes): intervaginal sponge of progesterone and PGF<sub>2α</sub> group (12 ewes): injection of PGF<sub>2α</sub>. The ewes P4 intervaginal sponges containing 40 mg of flugestone acetate (Syncrite-40, Australia), were left in the vagina for 12 days. In ewes of PGF<sub>2α</sub> were intramuscularly injected with two injection of PGF<sub>2α</sub> (1 ml Estromate, 250 mg Cloprostenol, Coopers Co., Germany) 12 days apart.

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Ewes were chocked for estrus signs for three times daily. Three fertile rams were introduced to the ewes in each group (one ram per 4 ewes) for estrus detection and mating; starting at the sponge withdrawal day in P4 group and after second injection of PGF<sub>2α</sub> in the morning and evening for 5 consecutive days. Estrus signs rate was calculated as the number of ewes showed signs of estrus per total ewes. The time of starting estrus was defined as the time between sponges removal or after second injection of PGF<sub>2α</sub> and the first accepted mount of the ewes was recorded. While in control group was recorded the time from 15 day after the last estrus and the first accepted mount of the ewes. Estrous duration was calculated as the time between the first and last accepted mount. Thirty days after successful last mating, ewes were examined for pregnancy by using ultrasound scanner (Honda HS-1500V, Honda Electronics, Japan). Conception rate calculated as the number of ewes conceived per initial number of ewes used. Gestation period was defined as the time between successful last mating and the parturition. The lambing rate was calculated as the number of ewes lambed per initial number of ewes used. Litter size was calculated as the number of total lambs born per number of ewe lambed. The body weights of lambs were weighed at birth and at weaning (120 days). The lambs were fasted overnight before weighing at 8.00 a.m. Weaning rate was calculated as number of lambs live at weaning per number of lambs born. Birth types (twins and single) and sex ratio (male and female) of lambs were recorded immediately after kidding.

Data were subjected to statistical analysis by using SPSS (2011) program. Differences among

treatment means were tested for significance ( $p < 0.05$ ) using Duncan's multiple range test (Duncan, 1955).

The mathematical model is:

$$Y_{ij} = \mu + T_i + e_{ij}$$

**Where:**

$Y_{ij}$  = Individual observation

$\mu$  = The overall mean for the trial under consideration

$T_i$  = The effect of the  $i$  treatments

$e_{ij}$  = Random residual error

## RESULTS

Estrus signs rate, the time of starting estrus, estrus duration, pregnancy rate, lambing rate, litter size and gestation length are shown in Table (1). The percentage of estrus signs exhibition in PGF<sub>2α</sub> group reached 83.3% ( $P < 0.05$ ) and in P4 group was 75%, while the lowest percentage (60%) was observed in control group. The time of starting estrus was significantly early ( $P < 0.05$ ) in treated groups than that of control group. The estrus duration was significantly longer ( $P < 0.05$ ) in treated groups than that of control group. The pregnancy and lambing rates were 100% in all ewes showed estrus in all groups, While pregnancy rate showed significant ( $P < 0.05$ ) differences between groups when calculated from the initial number used in each group. Estrus synchronization methods were significantly ( $P < 0.05$ ) raised the pregnancy rate in treated groups than that in non-treated (control group). The same trend and result was observed in lambing rate. There were no significant differences in the litter size among the three groups. It was 1.6 in P4 group, followed by 1.4 in PGF<sub>2α</sub> group and 1.3 in the control group. The gestation length showed about one day longer in P4 group than control and PGF<sub>2α</sub> groups.

**Table (1):** Estrus signs rate of ewes and reproductive parameters of treated and control groups (mean  $\pm$  SE)

Items	Control (10)	P4 (12)	PGF <sub>2α</sub> (12)
No. of ewes showed estrus	6	9	10
Estrus signs rate (%)	60 <sup>c</sup> (6/10)	75 <sup>b</sup> (9/12)	83.33 <sup>a</sup> (10/12)
Time of starting estrus (h)	58.40 <sup>a</sup> $\pm$ 2.85*	46.00 <sup>b</sup> $\pm$ 2.60	44.80 <sup>b</sup> $\pm$ 2.85
Estrus duration (h)	31.20 <sup>b</sup> $\pm$ 2.88	41.60 <sup>a</sup> $\pm$ 2.88	44.00 <sup>a</sup> $\pm$ 2.63
Pregnancy rate (%)	60 <sup>c</sup> (6/10)	75 <sup>b</sup> (9/12)	83.33 <sup>a</sup> (10/12)
Pregnancy of mated ewes (%)	100 (6/6)	100 (9/9)	100 (10/10)
Lambing rate (%)	60 <sup>c</sup> (6/10)	75 <sup>b</sup> (9/12)	83.33 <sup>a</sup> (10/12)
Lambd ewes of pregnant ewes (%)	100 (6/6)	100 (9/9)	100 (10/10)
Litter size	1.3 $\pm$ 0.15 (8/6)	1.6 $\pm$ 1.14 (14/9)	1.4 $\pm$ 0.14 (14/10)
Gestation length (days)	150.42 $\pm$ 0.50	151.83 $\pm$ 0.47	150.10 $\pm$ 0.42

\* The time from 15 day after the last estrus and the first accepted mount of the ewes  
<sup>a,b,c</sup> in the same row with different superscripts are significantly different ( $P < 0.05$ )

Table (2) shows birth weight, weaning weight, weaning rate, type of lambing of ewes and sex ratio of newborn lambs of control and estrus synchronized ewes. No significant differences in birth weight and weaning weight were found among the three groups. However, there was a trend of higher values in hormonally treated groups than that in control group on the previous parameters as (3.03 kg and 12.70 kg, respectively) in P4 group and (2.97 kg and 12.66 kg, respectively) in PGF<sub>2α</sub> group. Weaning rate was the highest (P<0.05) in PGF<sub>2α</sub> group (92.9%) and the lowest in P4 group (85.7%). The

percentage of ewes lambing twins was the highest (P<0.05) in P4 group (56%) and the lowest in control group (33%). Their counterparts of lambing single percentage was the highest (P<0.05) in control group (67%) and the lowest value in P4 group (44%). In addition, the percentage of male lambs was the highest (P<0.05) for ewes in PGF<sub>2α</sub> group (57.1%) and the lowest for ewes in control group (37.5%). Their counterparts of female lambs percentage was the highest (P<0.05) for ewes in control group (62.5%) and the lowest for ewes in PGF<sub>2α</sub> group (42.9%).

**Table (2):** Productive parameters of control and estrus synchronized ewes (mean ± SE)

Items	Control	P4	PGF <sub>2α</sub>
<b>Birth weight (kg)</b>	2.96 ± 0.08	3.03 ± 0.05	2.97 ± 0.05
<b>Weaning weight (kg)</b>	12.49 ± 0.25	12.70 ± 0.15	12.66 ± .015
<b>Weaning rate (%)</b>	87.5 <sup>b</sup> (7/8)	85.7 <sup>c</sup> (12/14)	92.9 <sup>a</sup> (13/14)
<b>Type of lambing:</b>			
<b>Single (%)</b>	67 <sup>a</sup> (4/6)	44 <sup>c</sup> (4/9)	60 <sup>b</sup> (6/10)
<b>Twins (%)</b>	33 <sup>c</sup> (2/6)	56 <sup>a</sup> (5/9)	40 <sup>b</sup> (4/10)
<b>Sex of newborn</b>			
<b>Male (%)</b>	37.5 <sup>c</sup> (3/8)	43 <sup>b</sup> (6/14)	57.1 <sup>a</sup> (8/14)
<b>Female (%)</b>	62.5 <sup>a</sup> (5/8)	57 <sup>b</sup> (8/14)	42.9 <sup>c</sup> (6/14)
<b>Sex ratio (female/male)</b>	1.7	1.3	0.75

<sup>a,b,c</sup> in the same row with different superscripts are significantly different (P<0.05)

## DISCUSSION

The current results showed successful estrus synchronization methods used in local ewes during summer season. The percentage of estrus signs rate exhibition in PGF<sub>2α</sub> group reached 83.3% and in P4 group was 75%, while it was 60% in control group. These findings were agreeable with Greyling and Van der Nest (2000), Motlomelo *et al.* (2002) and Naderipour *et al.* (2012) reported that the range of estrus signs rate in sheep was 51.7 to 87.5% using intravaginal progestagen sponges during the breeding and non-breeding season. Mutiga and Mukasa-Mugerwa (1992) and Ataman and Aköz (2006) found that estrus signs rate was 83% after double injection of PGF<sub>2α</sub>. Also, Abu El-Ella *et al.* (2016) found that the estrus signs rate was 86.7% in ewes treated by medroxyprogesterone acetate (MAP) + equine chorionic gonadotropin (eCG) and 60% in non-treated ewes during summer season. While the estrus signs rate determined in our study was little lower than that reported by Abdel-Mageed (2006), Akoz *et al.* (2006), Ozyurtlu *et al.* (2010), Pietroski *et al.* (2013) and Gardón *et al.* (2015) following sponge treatment. Öztürkler *et al.* (2003) and Abdalla *et al.* (2014) reported that estrus signs rate was 90.2 and 100% after double injection of PGF<sub>2α</sub>, respectively. Zeleke *et al.* (2005), Kareta *et al.* (2006), Kausar *et al.* (2009) and McCappin and Murray (2011) reported that variations in estrus signs rate could be attributed to age, parity, nutrition, breed, location and time of the year, climate changes and type of hormones. The time of

starting estrus varied between treatments and occurred within 24 - 144 h following progestagen or progesterone withdrawal (Simonetti *et al.*, 2000; Vinales *et al.*, 2001). In addition, Dogan and Nur (2006) recorded that ewes came to heat between 18 and 96 h after sponge withdrawal, with the highest incidence of estrus occurring between 30 and 60 h. The time of starting estrus in the present study lied through the previous reported research data. Moreover, our results showed that the time of starting estrus was earlier in P4 and PGF<sub>2α</sub> groups (46 and 44.8 h, respectively) than that in control group (58.4 h). The time of starting estrus in our study were lower than that reported by Didarkhah and Mesgaran (2013), Jackson *et al.* (2014) and Gardón *et al.* (2015) who found that the time of starting estrus was 60, 60 and 55 h in ewes treated with P4, respectively. In another study, Abu El-Ella *et al.* (2016) found that the time of starting estrus was 52.8 h using MAP + eCG during summer season. These results were higher than that reported by Almadaly *et al.* (2016) who found that time of starting estrus was 37.3 and 24 h with using P4 and PGF<sub>2α</sub>, respectively. Estrus duration (estrus period) was classified to be short (less than 25 h), normal (25-40 h) and long (more than 40 h) according to Deghady (2000). These differences may be explained by differences in breed, lactation, nutrition, season and gonadotropins or progestagen treatments (Romano, 2002 and Omontese *et al.*, 2010). In the present study, the estrus duration was significantly longer (44 h) in PGF<sub>2α</sub> and P4 groups (41.6 h) than that in control group

(31.2 h). Also, Abu El-Ella *et al.* (2016) found that estrus duration was 45.6 h, which was longer in hormonal treated ewes (MAP + eCG) than 24 h in the control group. Most of the recent developments in reproductive techniques have to use some hormones. Moreover, EL-Sherry *et al.* (2012) found that high pregnancy rates and litter size with using progesterone based in synchronized programs. Also, Yadi *et al.* (2011) showed that progestogens and PGF<sub>2α</sub> increased the conception rate in ewes. P4 and PGF<sub>2α</sub> either alone or in combination would be good techniques to improve reproductive performance and fertility (Loubsera and van Niekerka, 1981). All synchronization protocols had a positive effect on ewe fertility during the non-breeding season (Almadaly *et al.*, 2016). In our study, the pregnancy and lambing rates were 100% in all ewe came in estrus. This result agreed to results obtained by Jawad (2014) who found that the lambing rate of ewes treated by sponges was 100%. In the present study, calculating pregnancy rate values according to initial number of animals in each group showed that in PGF<sub>2α</sub> group (83%), in P4 group (75%) and in control group (60%). Almadaly *et al.* (2016) found that conception and lambing rates in ewes treated by P4 + eCG were 44% and 100% in ewes treated by PGF<sub>2α</sub>+ eCG. Gardón *et al.* (2015) found that pregnancy rate was 71.4% in ewes treated by MAP. Abdalla *et al.* (2014) found that the pregnancy and lambing rate were 95% in ewes treated by PGF<sub>2α</sub>. Also, Yadi *et al.* (2011) showed that pregnancy rate in ewes treated by sponges and PGF<sub>2α</sub> were 45 and 70%, respectively. On other studies, the conception rates in ewes treated by intra-vaginal sponges + eCG injected at time of sponge removal were 96.8% (Wildeus, 2000) and 100% (Huseyin and Yildiz, 2005; Akoz *et al.*, 2006). Also, the lambing rates were 85.7% (Akoz *et al.*, 2006), 75.6% (Koyuncu and Alticekic, 2010), 93.3% (Kulaksiz *et al.*, 2013) and 100% (Abdalla *et al.*, 2014; Abu El-Ella *et al.*, 2016). This discrepancy in the results reported by different researchers on pregnancy and lambing rate can be explained by the differences in body condition, breed, and management systems. Litter size is considered an important factor in the development of sheep production. Increasing litter size rates in sheep offers the best opportunity to increase the efficiency of lamb meat production. In our study, there were no significant differences in the liter size among the three groups. However, it was the highest (1.6) in P4 group and the lowest (1.3) in the control group. This result was similar to results obtained by Abdel-Megeed (2006) who found that litter size at birth was 1.3 in ewes treated with PGF<sub>2α</sub> and no significant differences between treated and not treated ewes. Also, Horoz *et al.* (2003) found that litter size at birth was 1.4 in ewes not hormonally treated. The litter size at birth in our study was higher than that reported by Gardón *et al.* (2015) who found that litter size was 1.4 in ewes treated by MAP sponges and Abu El-Ella *et al.* (2016) who found that litter size was 1.1 in not treated ewes. Safdarian *et al.* (2006) and Abdalla *et al.* (2014) found that litter size was 1.3 and 1.1 in ewes treated by PGF<sub>2α</sub>, respectively. Whereas, litter size was lower than that reported by Cruz *et al.* (1991) who found that litter size was 1.6 in ewes not

treated. In the present study, the gestation period showed about one day longer in P4 group (151.8 days) than control and PGF<sub>2α</sub> groups (150.4 days and 150.1 days, respectively). These results in agreement with those reported by Domingues *et al.* (1991) who found that the range of pregnancy period in ewes treated by sponges was reported from 144 days to 152 days. Also, Aboul-Ella (2006) reported that length of gestation period was shorter in ewes treated with PG in Barki ewes. Moreover, Zarkawi (2000) reported that the treatment had no effect on the duration of pregnancy, which averaged 150.3 days in control and 150.4 and 150.8 days in synchronized groups with 10 and 15 mg PGF<sub>2α</sub>, respectively. Whereas, pregnancy period in the present study was lower than that reported by Farrag (2019) who found that the average gestation length period were 157.37 and 154.88 days for control ewes and PGF<sub>2α</sub> ewes, respectively in Abou-Delik ewes grazing in the South Eastern zone of Egypt. Also, Safranski *et al.* (1992) who found that pregnancy periods in control ewes and ewes treated with melengesterol acetate (MGA) + PG-600 were 163.8 and 157.2 days, respectively in Kalkuhi ewes in Iran. Horoz *et al.* (2003) reported that gestation periods in control ewes and those treated with medroxyprogesteron were 164 days and 155 days, respectively in Kivircik ewes in Turkey. This difference can be likely due to the different breed of sheep and the environment. Body weight is considered to be a good indicator of the reproductive performance (Shetaewi *et al.*, 2001). Birth and weaning weights are important criteria in meat production because of their high correlation with growth rate and adult size (Al Shaikh *et al.*, 1989). In our study, no significant differences in birth weight and weaning weight among the three groups. However, it was the highest in P4 group and the lowest in the control group. These results were in agreement with those results obtained by Abu El-Ella *et al.* (2016) who found that birth weight was high in the treated groups than control group. Whereas, birth weight in present study was lower than that results reported by Abdel-Megeed (2006) which was 3.7 kg and Lethy *et al.* (2003) which was 3.5 kg in ewes treated by PGF<sub>2α</sub>. In present study weaning rate was significantly higher in PGF<sub>2α</sub> group than the other groups. These results were in agreement with result optioned by Farrag *et al.* (2010) who found that the weaning rate was 90% in ewes treated with two doses of PGF<sub>2α</sub>. In our study, the percentage of ewes lambled twins was the highest in P4 group (56%) and the lowest in control group (33%). This result was similar to that was found by Ezzat *et al.* (2017) who found that the incidence of twinning rate in ewe treated by 300 mg P4. Also, Abu El-Ella *et al.* (2016) found that the percentage of lambing single was higher in control than in MAP + eCG ewes.

## CONCLUSION

In general, using of P4 and PGF<sub>2α</sub> protocols were successful in inducing estrus synchronization of local ewes in terms of estrus signs rate, estrus duration, pregnancy and lambing rates compared to non-treated group. In addition, results indicated improving the

productive and reproductive performance of local sheep by applying estrus synchronization methods during summer season in Egypt.

### REFERENCES

- Abdalla, E. B., B. Farrag, A. L. S. Hashem, F. A. Khalil and M. S. Abdel-Fattah (2014). Effect of progesterone, PGF<sub>2α</sub>, PMSG AND GnRH on estrus synchronization and some reproductive and productive traits in Barki ewes. *J. of Agroalimentary Processes and Techno.*, 20(1): 93-101.
- Abdel-Megeed, I. I. (2006). Economic evaluation of lamb production in Rahmani sheep under two regimes for super-ovulation, Ph.D. Thesis, Fac. Agric, Cairo. Univ.
- Abecia, J. A., F. Forcada and A. González-Bulnes (2012). Hormonal control of reproduction in small ruminants. *Anim. Reprod. Sci.*, 130: 173-179.
- Aboul-Ella, A. A. (2006). Rate of Barki ewes to treatment with gonadotrophin hormones and energy supplementation (flushing). *Egyptian Journal of Sheep, Goat and Desert Animals Sciences*, 1(1): 73-88.
- Abu EL-Ella, A. A., D. F. Teleb, M. A. M. Abdel-Hafez and A. M. Deghedy (2016). Appraisal of different protocols for estrus synchronization in local Rahmani sheep. *Egyptian J. of Sheep and Goat Science*, 11(3): 116-131.
- Akoz, M., B. Bulbul, M. B. Ataman and S. Dere (2006). Induction of multiple births in Akkaraman cross-bred sheep synchronized with short duration and different doses of progesterone treatment combined with PMSG outside the breeding season. *Bull. Vet. Inst. Pulawy*, 50: 97-100.
- Almadaly, E., M. Ashour, I. El-Kon, B. Heleil and E. Fattouh (2016). Efficacy of various synchronization protocols on the estrus behavior, lambing rate and prolificacy in Rahmani Egyptian ewes during the non-breeding season. *Asian J. of Anim. and Veter. Advances*, 11: 34-43.
- Al Shaikh, M. A., H. H. Mogawer, M. S. Salah, M. N. Bakkar and H. H. Mogawer (1989). Body weight of Aradi goat kids in Saudi Arabia at different ages and affecting factors. *J. King Saud Univ. I, Agric. Sci.*, 1: 17-24.
- Ataman, M.B. and M. Akoz (2006). GnRH-PGF<sub>2α</sub> and PGF<sub>2α</sub>-PGF<sub>2α</sub> synchronization in Akkaraman cross-bred sheep in the breeding season. *Bull. Vet. Inst. Pulawy.*, 50: 101-104.
- Ataman, M. B., M. Akoz and O. Akman (2005). Induction of synchronized oestrus in Akkaraman cross-bred ewes during and outside the breeding season: Use of short-term and long-term progesterone treatments. *Proceedings of the International 6th Sheep Veterinary Congress, Crete, Greece*, 130-131.
- Cruz, D. G., M. J. De La Castaneda and C. G. Rocha (1991). Effects of estrus synchronization by means of FGA-impregnated sponges on the fertility and prolificacy of partly housed pelibuey ewes. *Anim Breed Abstr.*, 10: 52-59.
- Davis, A. J., I. R. Fleet, F. A. Harrison and F. M. M. Walker (1980). Pulmonary metabolism of prostaglandin F<sub>2α</sub> in the conscious non-pregnant ewe and sow. *J. Physiol.*, 301: 86-86.
- Deghady, A. M. (2000). Seasonal elements affecting reproduction of Rahmani ewes. M.Sc. Thesis Fac. of Agric. Menoufia University.
- Didarkhah, M. and M. D. Mesgaran (2013). Comparison of two methods estrus synchronization by CIDR and sponge along with PMSG various levels on Baloochi ewes on reproductive performance in breeding season. *J. Am. Sci.*, 9: 168-172.
- Dogan, I. and Z. Nur (2006). Different estrous induction methods during the non-breeding season in Kivircik ewes. *Vet. Med-Czech*, 51: 133-138.
- Domingues, F., J. C. dez-Tejerina., J. Miro-Roig and M. Carbajo Rudea (1991). Induction and synchronization of estrus during seasonal anestrus in improved Ripollesa ewes by means of FGA-impregnated vaginal sponges and PMSG injections. *Anim Breed Abstr.*, 59: 1797.
- Duncan, D. B. (1955). Multiple range and multiple F tests. *Biometrics*, 11: 1-42.
- EL-Sherry, T. M., D. R. Derar, M. Hayder, H. Hamdon and E. Saifelnasr (2012). Effect of progesterone and prostaglandin analogue-based synchronization programs on the follicular dynamics and conception rate at two different breeding seasons in subtropical ewes. *Assiut Vet. Med. J.*, 58(135): 88-96.
- Ezzat, A. A., A. S. El-Naeim and I. K. Sobhy (2017). CIDR and Ultrasound-Fetometry Scan; Comparative Study Between Sheep and Goat in Egypt. *AJVS*. 53: 46-56.
- Farrag, B. (2019). Productive Characteristics and Reproductive Rates to Estrus Synchronization and Flushing in Abou-Delik Ewes Grazing in Arid Rangelands in Halaieab - Shalateen - Abouramad Triangle of Egypt. *World Vet. J.*, 9(3): 201-210.
- Farrag, B., A. L. S. Hashem, M. T. Badawy, E. E. Tharwat and F. A. Khalil (2010). Estrus synchronization of Barki ewes using prostaglandin F<sub>2α</sub> and their rate to different doses of PMSG. *Research Bulletin, Ain Shams Univ.*, 1-17.
- Gardón, J. C., B. Escribano, S. Astiz and S. Ruiz (2015). Synchronization protocols in Spanish Merino Sheep: reduction in time to estrus by the addition of ecG to a progesterone-based estrus Synchronization protocol. *Anim. Sci.*, 15(2): 409-418.
- Gibson, W. R. and T. J. Robinson (1971). The seasonal nature of reproductive phenomena in the sheep. I. Variation in sensitivity to oestrogen. *J. Reprod. Fertil.*, 24: 9-18.

- Greyling, J. P. C. and M. Van der Nest (2000). Synchronization of oestrus in goats: dose effect of progestagen. *Small Ruminant Research*, 36: 201-207.
- Horoz, H., G. Kasikci, K. Ak, S. Aklan and C. Sonmez (2003). Controlling the breeding season using melatonin and progestagen in Kivircik ewes. *Turk J Vet Anim Sci*, 27: 301-305.
- Husein, M. Q., M. M. Ababneh and S. G. Haddad (2005). The effects of progesterone priming on reproductive performance of GnRH-PGF2a-treated anestrus goats. *Reprod. Nutr. Dev.* 45: 689-698.
- Husein, M. Q. and R.T. Kridli (2003). Effect of progesterone prior to GnRH-PGF2a treatment on induction of oestrus and pregnancy in anoestrus Awassi ewes. *Reprod. Dom. Anim.*, 38: 228-232.
- Huseyin, T. and H. Yildiz (2005). Synchronization of oestrus in Hamdani ewes: The use of different PMSG doses. *Bull. Vet. Inst. Pulawy*, 49: 311-314.
- Huston, J. E. (1983). Production of fine-wool ewes on yearlong rangeland in West Texas. II. Effects
- Jackson, C. G., T. L. Neville, V. R. G. Mercadante, K. M. Waters and G. C. Lamb (2014). Efficacy of various five-day estrous synchronization protocols. *Sm. Rumin. Res.*, 120: 100-107.
- Jawad, K. Taher (2014). Different oestrus induction methods in awassi ewes during the out of breeding season. *Bas. J. Vet. Res.*, 1(2): 66-74.
- Kareta, W., K. Korman and M. Cegla (2006). Ovulation level and prolificacy in ewes depending on their age, birth type and percentage of prolific genotype. *Reprod. Biol.*, 2: 73-78.
- Kausar, R., S. A. Khanum, M. Hussain and M. S. Shah (2009). Estrus synchronization with medroxyprogesterone acetate impregnated sponges in goats (*capra hircus*) Pakistan *Vet. J.*, 29(1): 16-18.
- Khalifa, R. M. (1993). None surgical intrauterine artificial insemination in sheep using exogenous oxytocin. *Egypt J. Anim. Prod.*, 30(1): 55-61.
- Knights, M., Q. S. Baptiste and P. E. Lewis (2002). Reproductive Performance of anestrus ewes treated with used-CIDR devices and estrogen. Joint Annual Meeting of the ADSA/ASAS Quebec City, Canada, July, 21-25. *J. Anim. Sci.*, 80: (Suppl. 1): 297.
- Koyuncu, M. and S. O. Alticekic (2010). Effects of progestagen and PMSG on estrous synchronization and fertility in Kivircik ewes during natural breeding season *Asian-Aust. J. Anim. Sci.*, 23(3), 308-311.
- Kulaksiz, R., O. Ucar and A. Daskin (2013). Effects of FGA sponge and ovsynch based protocols on reproductive performance of fat-tailed ewes during the breeding season. *Kafkas Univ. Vet. Fak. Derg.* 19(4): 629-633.
- Lethy, H. M., O. A. Salama and E. E. Tharwat (2003). Estrus synchronization and artificial insemination in Rahmani ewes: A comparison of two methods of insemination. *Annals Agric. Sci. Ain Shams Univ.*, 48(1): 107-116.
- Loubsera, P. and C. van Niekerka (1981). Oestrus synchronization in sheep with progesterone-impregnated (MAP) intravaginal sponges and a prostaglandin analogue. *Theriogenology*, 15: 547-552.
- McCappin, N. and R. D. Murray (2011). Factors affecting the pregnancy rate in ewes following AI. *Vet. Rec.*, 168: 99.
- Motlomelo, K. C., J. P. C. Greyling and L. M. J. Schwalbach (2002). Synchronization of oestrus in goats: the use of different progestagen treatments. *Small Rumin. Res.*, 45: 45-49.
- Mutiga, E. R. and E. Mukasa-Mugerwa (1992). Effect of the method of estrus synchronization and PMSG dosage on estrus and twinning in Ethiopian Menze sheep. *Theriogenology*, 38: 727-734.
- Naderipour, H., J. Yadi, A. G. Shad and M. A. Sirjani (2012). The effects of three methods of synchronization on estrus induction and hormonal profile in Kalkuhi ewes: A comparison study. *African J. of Biotech*, 11(5): 530-533.
- NRC (1985). National Research Council, Nutrient Requirements of Domestic Animals. Nutrient requirements of sheep. National Academic Press. Washington, D.C. USA.
- Oliveira, M. E. F., H. Ayres, L.G. Oliveira, F.F.P.C. Barros and E. Oba (2015). Effects of season and ovarian status on the outcome of long-term progesterone-based estrus synchronization protocols and ovulatory follicle development in Santa Ines ewes under subtropical conditions. *Theriogenology*, 10: 9-24.
- Omontese, B. O., P. I. Rekwot, H. J. Makun, J. A. Obidi, J. S. Ruwaan and N. P. Chiezey (2010). Synchronization of estrus using EAZI Breed TM CIDR and FGA-30 intravaginal sponge in pre-partum Yankasa ewes. *Res. J. of Anim. Sci.*, 4(1): 53-57.
- Öztürkler, Y., A. Baykal and B. G. Ven (2003). Combined effect of a prostaglandin analogue and a progestagen treatment for 5 days on oestrus synchronisation in Tushin ewes. *Indian Vet. J.*, 80: 917-920.
- Ozyurtlu, N., I. Kucukaslan and Y. Cetin (2010). Characterization of oestrus induction rate, oestrus duration, fecundity and fertility in Awassi ewes during the non-breeding season utilizing both CIDR and intravaginal sponge treatments. *Reprod Domest Anim.*, 4(5): 464-467.
- Panhwar, F. (2007). Modern reproductive methods used to enhance goat production. Agricultural Research Services, United States Department of Agriculture, 1999-2008.

- Pietroski, A. C. C. A., F. Z. Brandão, J. M. G. de Souza and J. F. de Fonseca (2013). Short, medium or long-term hormonal treatments for induction of synchronized estrus and ovulation in Saanen goats during the nonbreeding season. *Rev Bras Zootec*, 42: 168-173.
- Romano, J. E. (2002). Does in proestrus-estrus hasten estrus onset in does estrus synchronization during the breeding season. *Applied Anim. Behav. Sci.*, 77: 329-334.
- Safdarian, M., M. Kafi and M. Hashemi (2006). Reproductive performance of Karakul ewes following different oestrous synchronisation treatments outside the natural breeding season. *South African Journal of Animal Science*, 36(4): 229-234.
- Safranski, T. J., W. R. Lamberson and D. H. Keisler (1992). Use of melengesterol acetate and gonadotropins to induce fertile estrus in seasonally anestrous ewes. *J. Anim. Sci.*, 70: 2935-2941.
- Shetaewi, M. M., A. M. Abdel-Samee and E.A. Bakr (2001). Reproductive performance and milk production of Damascus goats fed acacia shrubs or berseem clover hay in North Sinai, Egypt. *Tropical Animal Health and Production*, 33(1): 67-79.
- Simonetti, L., G. Ramos and J. C. Gardon (2002). Effect of estrus synchronization and artificial insemination on reproductive performance of Merino sheep. *Braz. J. Vet. Res. Anim. Sci.*, 39: 143-146.
- Simonetti, L., M. R. Blanco and J. C. Gardon (2000). Estrus synchronization in ewes treated with sponges impregnated with different doses of medroxyprogesterone acetate. *Small Ruminant Research*, 38: 243-247.
- SPSS, (2011). *SPSS Statistics for Windows, Version 20.0*. Armonk, NY, USA: IBM Corp.
- Turk, G., S. Gur, M. Sonmez, T. Bozkurt, E. H. Aksu and H. Aksoy (2008). Effect of exogenous GnRH at the time of artificial insemination on reproductive performance of Awassi ewes synchronized with progestagen-PMSG-PGF<sub>2α</sub> combination. *Reprod. Domest. Anim.*, 43: 308-313.
- Vinoles, C., B. Paganoni, J. T. B. Milton, M. A. Driancourt and G. B. Martin (2011). Pregnancy rate and prolificacy after artificial insemination in ewes following synchronisation with prostaglandin, sponges, or sponges with bactericide. *Anim. Prod. Sci.*, 51: 565-569.
- Vinoles, C., M. Forsberg, G. Banchemo and E. Rubianes (2001). Effect of long-term and short-term progestagen treatment on follicular development and pregnancy rate in cyclic ewes. *Theriogenology*, 55: 993-1004.
- Wildeus, S. (2000). Current concepts in synchronization of estrus: Sheep and goats. *J. Anim. Sci.*, 77: 1-14.
- Yadi, J., M. F. Moghaddam, S. Khalajzadeh and A. A. Solati (2011). Comparison of Estrus Synchronization by PGF<sub>2α</sub>, CIDR and Sponge with PMSG in Kalkuhi Ewes on Early Anestrous Season. *IPCBE*, 13:61-65.
- Zarkawi, M. (2000). The evaluation of two doses of prostaglandin F<sub>2α</sub> analogue, prosolvin, for oestrous synchronization of Syrian Awassi ewes. *New Zealand Journal of Agricultural Research*, 43: 509-514.
- Zelege, M., J. P. C. Greyling, L. M. J. Schwalbach, T. Muller and J. A. Erasmus (2005). Effects of progestagen and PMSG on oestrus synchronization and fertility in Dorper ewes during the transition period. *Small Ruminant Res.*, 56: 47-53.

## استخدام البروجسترون والبروستاجلاندين $F_{2\alpha}$ في تزامن الشياح في الأغنام المحلية خلال فصل الصيف في مصر

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أجريت هذه الدراسة لمعرفة تأثير استخدام هرمونات البروجسترون والبروستاجلاندين  $F_{2\alpha}$  على تزامن الشياح في فصل الصيف في مصر، ومقارنة استخدام هذه الهرمونات في إحداه تزامن الشياح مع الشبق الطبيعي في النعاج. تم استخدام أربعة وثلاثين نعجة محلية غير مرضعة وغير عشار أعمارهم ما بين ٢-٤ سنة وبمتوسط وزن حى  $١.٨٠ \pm ٤٢.١$  كجم. تم تقسيم النعاج بشكل عشوائي إلى ثلاث مجموعات: مجموعة الشياح الطبيعي (المجموعة غير المعاملة،  $n = 10$ )، مجموعة الإسفنجات ( $P4=12$ ) لمدة ١٢ يوماً ومجموعة البروستاجلاندين  $F_{2\alpha}$  (١٢ نعجة) بمعدل حقنيتين بفارق زمني ١٢ يوماً. تم قياس معدلات الشبق والحمل وأوزان المواليد. أظهرت النتائج أن نسبة الاستجابة للشبق في النعاج المعاملة البروستاجلاندين  $F_{2\alpha}$  بلغت ٨٣.٣% في حين لوحظ أن أقل نسبة (٦٠%) في المجموعة غير المعاملة ( $P < 0.05$ ). وكانت أقصر فترة للشياح المتوقع ( $P < 0.05$ ) في مجموعات P4 والبروستاجلاندين  $F_{2\alpha}$  (٤٦ و ٤٤,٨ ساعة على التوالي) عن المجموعة غير المعاملة (٥٨,٤ ساعة). في حين كانت مدة الشبق أطول معنويًا ( $P < 0.05$ ) في مجموعات البروستاجلاندين  $F_{2\alpha}$  و P4 (٤٤ و ٤١,٦ ساعة على التوالي) عن المجموعة غير المعاملة (٣١,٢ ساعة). وكانت معدلات الحمل والولادة ١٠٠% في جميع المجموعات. بينما كانت معدلات الحمل والولادة المنسوبة إلى الأرقام الأولية المستخدمة ٨٣% و ٧٥% و ٦٠% للمجموعات البروستاجلاندين  $F_{2\alpha}$  و P4 والمجموعة غير المعاملة ( $P < 0.05$ ) على التوالي. بينما لا توجد اختلافات معنوية بين المجموعات في معدل المواليد والوزن عند الولادة ووزن الفطام. في حين كان معدل الفطام أعلى معنويًا ( $P < 0.05$ ) في مجموعة البروستاجلاندين  $F_{2\alpha}$  (٩٢,٩%) عن المجموعة غير المعاملة ومجموعة P4 (٨٧,٥ و ٨٥,٧% على التوالي). وكذلك كانت أعلى معدل التوائم (٥٦%) في مجموعة البروجسترون وأقل معدل التوائم (٣٣%) في مجموعة المجموعة غير المعاملة. وأيضاً كانت أعلى نسبة للذكور المولودة ( $P < 0.05$ ) في مجموعة البروستاجلاندين  $F_{2\alpha}$  (٥٧,١%) وأقل نسبة للذكور المولودة في المجموعة غير المعاملة (٣٧,٥%). ومما سبق فإن الدراسة أوضحت أن استخدام البروستاجلاندين  $F_{2\alpha}$  والبروجسترون يمكن أن يحسنوا في تزامن الشبق وبعض الصفات التناسلية والإنتاجية في النعاج المحلية خلال موسم الصيف في مصر. ويوصى بإجراء مزيد من الدراسات لإثبات النتائج باستخدام عدد أكبر من النعاج.